

Baicalein induces apoptosis and autophagy of breast cancer cells via inhibiting PI3K/AKT pathway in vivo and vitro

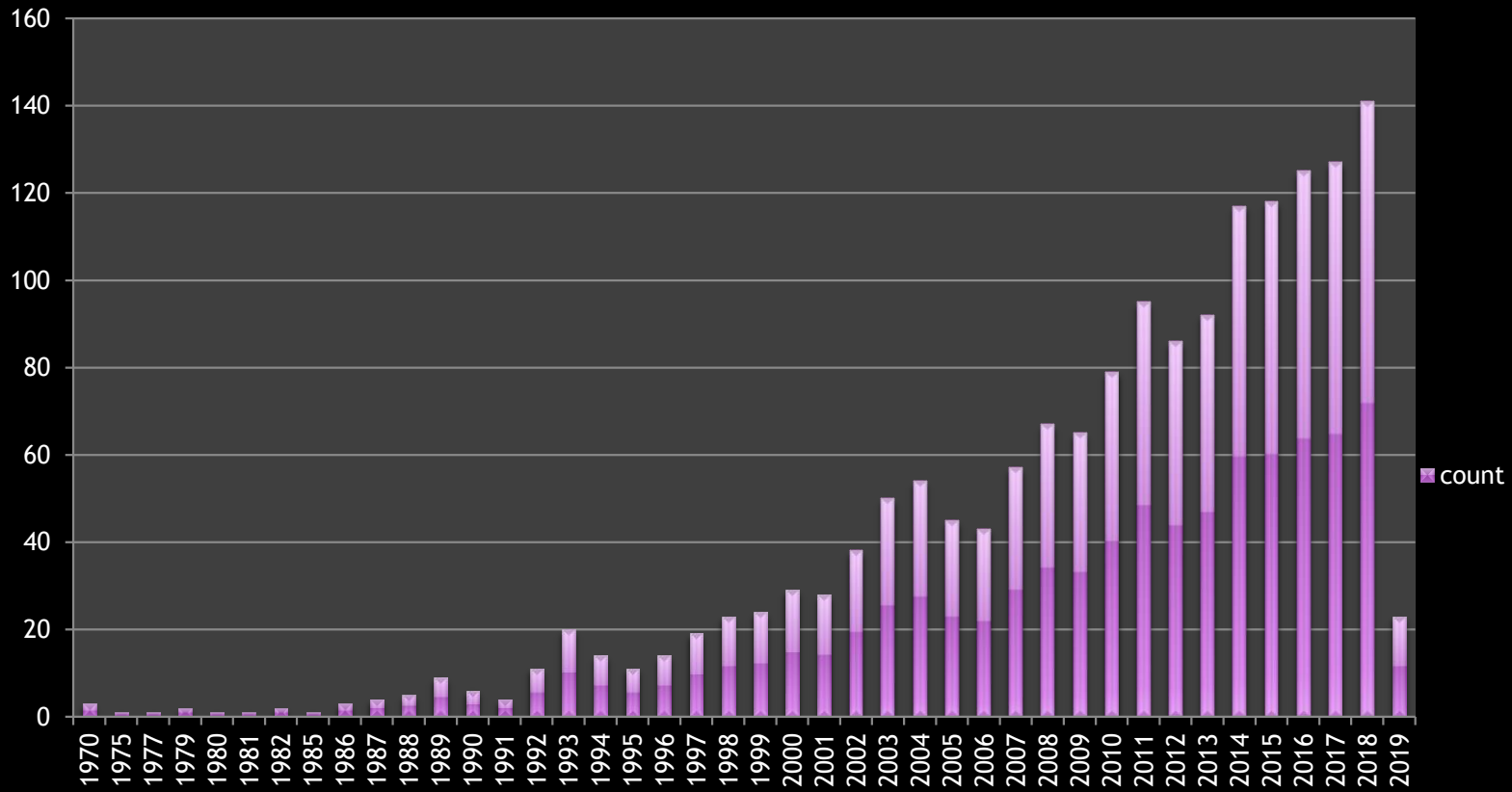
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“List of Contents”

- ◉ **Journal information**
- ◉ **Introduction**
- ◉ **Materials & methods**
- ◉ **Results**
- ◉ **Discussion**
- ◉ **Conclusion**

Journal information

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Introduction

❖ Breast cancer:

- ⊙ One of the most commonly occurring female malignant tumors.
- ⊙ Increased incidence and much **younger** onset age recently.
- ⊙ Although there have been significant advances in screening ,surgery, and chemo radiotherapy techniques, the **prognosis** of patients remains little known.
- ⊙ Hence, it is urgent to provide a new therapeutic strategy in cancer therapy.

Introduction

❖ Baicalein:

- ⊙ A bioactive component extracted from the root of *Scutellaria baicalensis* Georgi.
- ⊙ **Anti-tumor, anti-inflammatory, anti-cardiovascular disease, & antimicrobial** activities
- ⊙ Numerous studies: anti-tumor properties of baicalein in many types of human cancer cell lines both in vitro & in vivo.
- ⊙ Inhibiting cell **proliferation**.
- ⊙ Inducing cell **apoptosis** (activating the caspase cascade & mitochondrial apoptotic pathway, DNA fragmentation in malignant cells).

Introduction

- ◉ Past studies: Baicalein-induced apoptosis & autophagy via PI3K/AKT signaling pathway inhibition in renal carcinoma, glioma, epidermoid carcinoma, and bladder cancer.
- ◉ Few studies: were on latent molecular mechanism of anticancer activity of baicalein on human breast cancer cells.
- ◉ Present study purpose: ascertain potential mechanisms through which baicalein induces apoptosis & autophagy in MCF7 and MDA-MB-231 cells: (PI3K/AKT inhibition).
- ❖ PI3K/AKT signaling pathway play roles in :
 - ◉ Mammalian cell proliferation, differentiation, apoptosis, autophagy , & survival.
 - ◉ Survival of human cancer cells in vitro.
 - ◉ Carcinogenicity, invasion and metastasis of human cancer cells in vivo.

Materials & methods

- ◉ Cell culture
- ◉ MCF-7 and MDA-MB-231 cells: were cultured in **DMEM** supplemented with 10% **FBS**, 50 IU/mL **penicillin**, and 50 µg/mL **streptomycin** and maintained in humidified **5%** CO₂ air at **37°C**.
- ◉ Animals:
- ◉ Female **BALB/c** nude mice (3–6 weeks old, body weight 18–20 g) (from Experimental Animal Center of Xi'an Jiaotong University).

Materials & methods

Morphological cell changes

- ⦿ Cells were seeded in 12-well plates at a density of 3×10^5 cells/well and grown for 24 hours.
- ⦿ Cells were treated with various concentrations (0, 10, 20, and 40 μM) of baicalein.
- ⦿ grown at 37°C, in 5% CO₂ and 95% air for 24 hours.
- ⦿ For examining morphological changes, cells were observed and photographed under a phase-contrast microscope.

Materials & methods

MTT assay

- ◉ The cell viability was assessed by MTT assay.
- ◉ Cells were **seeded** into 96-well plates (4×10^3 cells/well) .
- ◉ **Incubated** in 5% CO₂ air at 37°C.
- ◉ After 12h incubation, the cells were **treated** with different concentrations of baicalein (0, 10, 20, and 40 μ M) for 24, 48, and 72 hours.
- ◉ **MTT** (20 μ L of 5 mg/mL) was added to each well and **incubated** at 37°C for 4 hours.
- ◉ The formazan crystals were dissolved in **DMSO**, then incubated for 10 min.
- ◉ The **OD** was recorded at 490 nm on a microplate reader.
- ◉ The inhibition ratio (IR) = (1-mean OD value of experimental group/mean OD value of control group) \times 100%.

Materials & methods

Colony formation assay

- ◉ Cells (1000 cells/well) were **seeded** into 6-well plates.
- ◉ **Treated** with baicalein (0, 10, 20, and 40 μ M) for 48 hours.
- ◉ Then cultured in DMEM at 37°C for **14 days**.
- ◉ During this period, the cells were **washed** with PBS every 3 days.
- ◉ **Fixed** with paraformaldehyde (4%, 15 minutes)
- ◉ **Stained** with crystal violet (0.1%, 15 minutes) and the numbers of colonies with >50 cells were **counted** with an Olympus digital camera.

Materials & methods

Hoechst 33258 staining for apoptosis

- ◉ Cells (2×10^5 cells/well) were **seeded** in 12-well plates for 12 hours
- ◉ **Treated** with various concentrations of baicalein (0, 10, 20, and 40 μM) for 48 hours
- ◉ **Washed** with PBS, and then **fixed** in paraformaldehyde (4%) for 15 minutes at room temperature.
- ◉ Cells were **stained** with 100 μL Hoechst 33258 in PBS for 15 minutes at room temperature.
- ◉ The stained cells were visualized from randomly selected fields under a fluorescence **microscope** (The nuclear condensation and fragmentation of cells were identified as the apoptotic cells).

Materials & methods

Measurement of mitochondrial membrane potential ($\Delta\Psi_m$)

- Cells were **seeded** in 12-well plates at a density of 2×10^5 cells/well for 12 hours.
- **Treated** with different concentrations of baicalein (0, 10, 20, and 40 μM) for 48h.
- Cells were **harvested** and **washed** with PBS and **resuspended** in JC-1 at 37°C for 30 minutes in the dark.
- The **stained** cells were analyzed by a fluorescence microscope.

Materials & methods

AO staining

- ⊙ MCF-7 and MDA-MB-231 cells (1×10^5 cells/well) were suspended and **seeded** in a 12-well plate and incubated overnight.
- ⊙ **Treated** with baicalein (0, 10, 20, and 40 μM) for 48 hours.
- ⊙ **Stained** with AO (1 $\mu\text{g/mL}$) for 15 minutes in the dark
- ⊙ **Washed** with PBS, and **visualized** under a fluorescence microscope.

Materials & methods

Transmission electron microscopy (TEM) observation (to observe the ultrastructure)

- ◉ Cells were incubated with 40 µg/mL baicalein for 48 hours.
- ◉ Then cells were **fixed** in ice-cold glutaraldehyde in 0.1 mol/L phosphate buffer (PH 7.4) overnight.
- ◉ **Fixed** in 1% osmium tetroxide and dehydrated.
- ◉ Cells were impregnated with **Epon**.
- ◉ The ultrathin sections were contrasted with **uranyl acetate** and **lead citrate** for electron microscopy.
- ◉ Electron micrographs were **observed** through a transmission electron microscope.

Materials & methods

Flowcytometry analysis

- ⊙ **Treatment** with various concentrations (0, 10, 20, and 40 μM) of baicalein for 48h.
- ⊙ Cells were collected and **washed** twice in PBS at room temp.
- ⊙ Cells (1×10^6) were **resuspended** in 100 μL annexin-V binding buffer solution.
- ⊙ 5 μL **annexin V-FITC** and 5 μL **propidium iodide** (PI) were added.
- ⊙ **Incubated** at room temp. for 15 min in the dark.
- ⊙ The percentage of apoptotic cells was analyzed by flowcytometry.

Materials & methods

Quantitative real-time PCR(qRT-PCR)

- ◉ Cells were **treated** with various concentrations (0, 10, 20, and 40 μ M) of baicalein for 48 hours.
- ◉ Total cell **RNA was extracted** by Trizol according to the manufacturer's instructions.
- ◉ RNA was then reverse **transcribed into cDNA** with RevertAid First-Strand cDNA Synthesis Kit.
- ◉ Then qRT-PCR reaction system was prepared following the manufacturer's instructions given in the SYBR®Premix Ex TaqII RT-PCR Kit (TaKaRa) using 100 ng cDNA.
- ◉ Each sample was analyzed by using a StepOnePlus Real-Time PCR System (Thermo Fisher Scientific).

Materials & methods

Table 1 Primers used for qRT-PCR analysis

Gene	Primer sequence	
	Forward (5'-3')	Reverse (5'-3')
Bcl-2	TCGCCCTGTGGATGACTGA	CAGAGACAGCCA GGAGAAATCA
Bax	GCGATGAACTGGACAACA ACAT	TAGCAAAGTAGAAAAGGGCAACC
LC3	GAGTGGAAGATGTCCGGCTC	CCAGGAGGAAGAAGGCTTGG
BECN1	CGGGATCCATGGAAGGGTCTAAGACGTCC	CGGAATTCTCATTTGTTATAAAATTGTGAGG
GAPDH	ATGCCAGTGAGCTTCCCGTCAGC	GGTATCGTGGAAGAACTCATGAC

Abbreviation: qRT-PCR, quantitative real-time PCR.

Materials & methods

Western blotting

- ⊙ Cells **cultured** with various concentrations (0, 10, 20, and 40 μ M) of baicalein for 24 hours, 48 hours, and 72 hours.
- ⊙ Proteins were separated by 10% **SDS-PAGE** and transferred onto PVDF **membranes**.
- ⊙ Membranes were **blocked** with 5% non-fat milk for 2h at room temp. & incubated overnight at 4°C with a **primary Ab** (GAPDH, AKT, p-AKT, mTOR, p-mTOR, NF- κ B, I κ B, and p-I κ B).
- ⊙ Membranes were **washed** 3times with TBST & incubated with the appropriate HRP-conjugated **secondary Ab** for 2h at room temp.
- ⊙ Protein bands were **visualized** using the chemiluminescence gel imaging system & quantitated using the Image-Pro Plus 6.0 software (GAPDH=internal control).

Materials & methods

Tumor xenograft study

- ◉ Female BALB/c nude mice (3–6 weeks old).
- ◉ Xenografts were established by subcutaneous injection of MCF-7 ($1 \times 10^7/200 \mu\text{L}$) & MDA-MB-231 cells (2×10^6 cells/ $200 \mu\text{L}$) into the 2left breast pad of nude mice.
- ◉ When tumor size $\cong 100 \text{ mm}^3 \rightarrow$ mice were randomly divided into different groups (5 in each) and baicalein (100 mg/kg) or 1% carboxymethyl cellulose sodium was administered by intra-gastric gavage 1/day for 21 days.
- ◉ Tumor **volume** , **growth** and body **weight** were measured every 3days.
- ◉ Experiments were terminated at the 21st day and the animals were anesthetized and sacrificed. Tumors were removed for further analysis.

Materials & methods

Immunohistochemistry

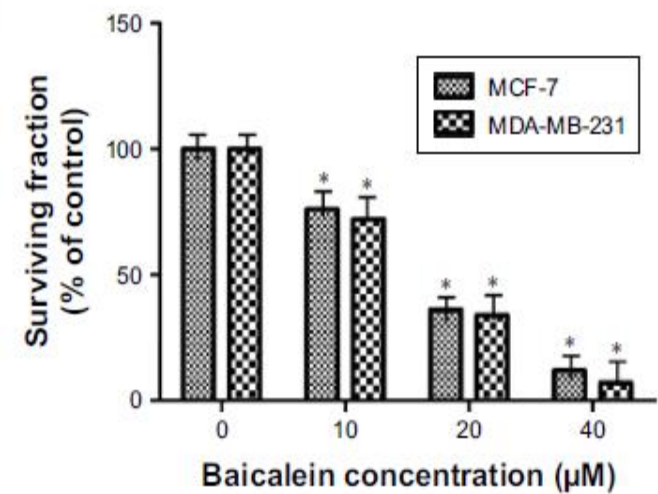
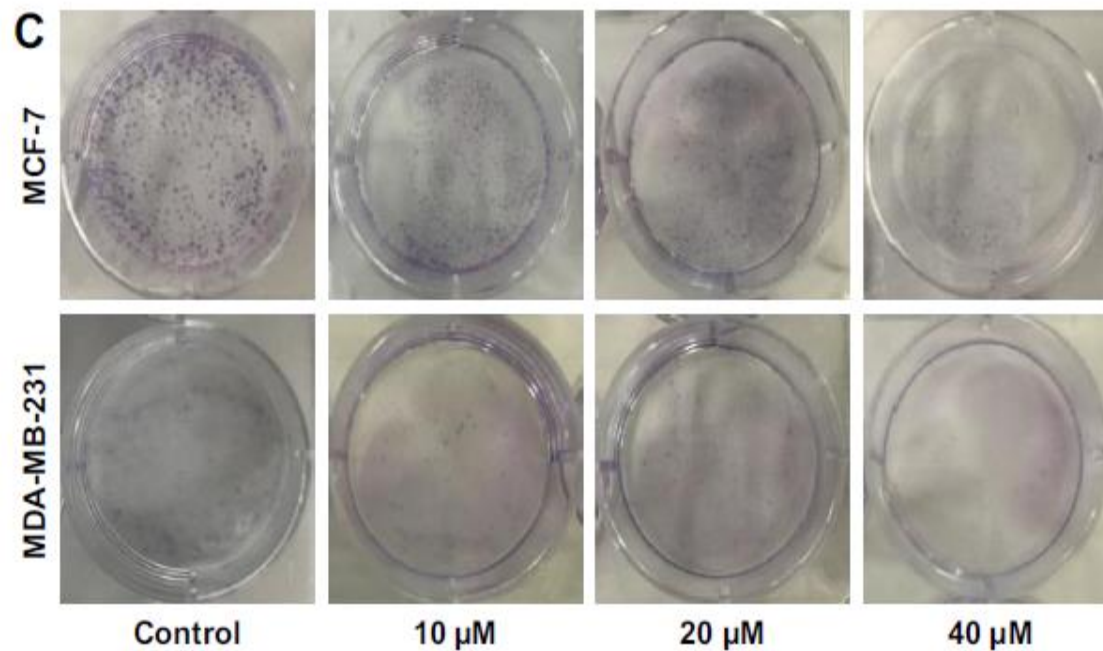
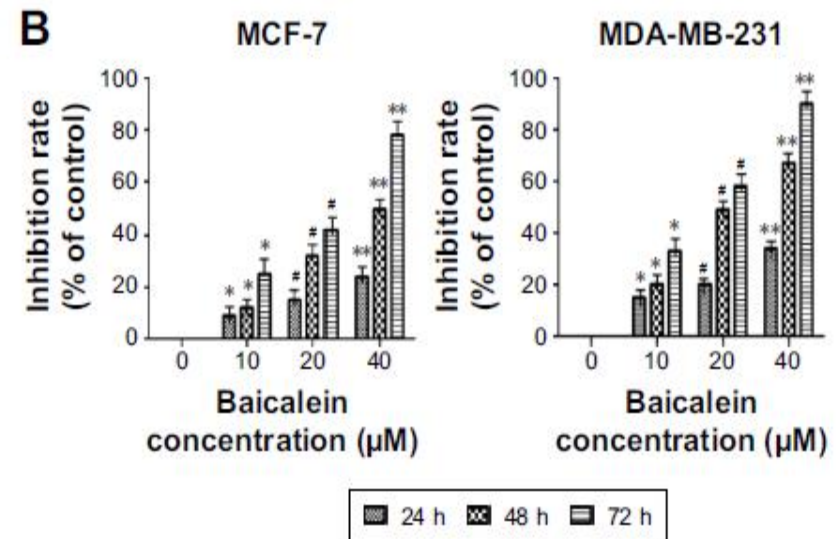
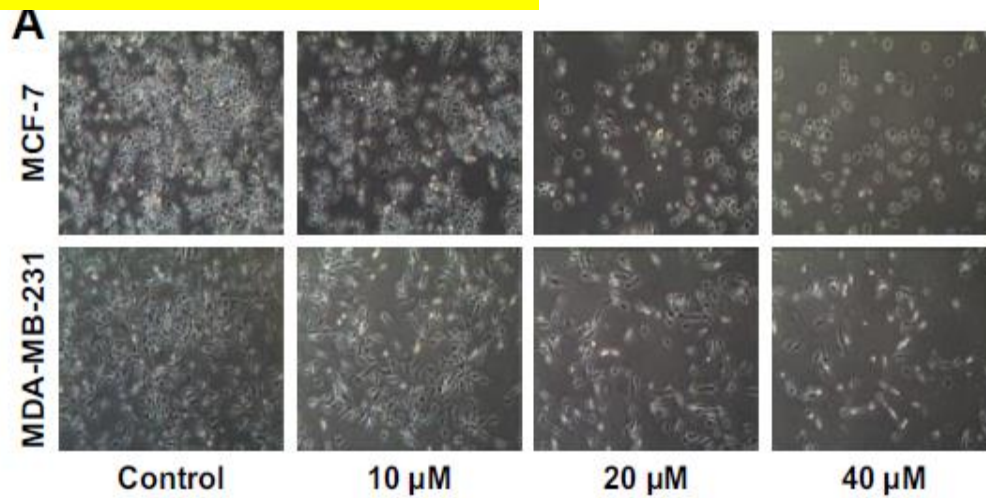
- ◉ Histological analysis was performed on tissue samples isolated from mouse xenografts.
- ◉ Briefly, 5 μ m sections were cut from all paraffin blocks and stained for p-AKT, Bax, and LC3.
- ◉ Immunohistochemical **staining** was conducted using Histostain®-SP Kits according to the manufacturer's instructions.
- ◉ Sections were incubated with p-AKT, BAX, and LC3 **primary antibody** at 4°C overnight.
- ◉ Images of sections were **visualized** using a Zeiss microscope & analyzed by Image-Pro Plus 6.0.

Results

Baicalein inhibited the **proliferation** of breast cancer cells

- ❖ **Phase-contrast Microscopy:**
 - ⊙ MCF-7&MDA-MB-231 cells exposed to increasing concentrations of baicalein → significant morphological changes, including cell shrinkage and blebbing (Fig.1A).
 - ⊙ MTT assay: baicalein significantly inhibited the proliferation of MCF-7 and MDA-MB-231 cells in a dose- and time-dependent manner (Fig.1B)
 - ⊙ Baicalein suppressed the colony formation of MCF-7 and MDA-MB-231 cells as shown by the plate colony formation assay (Fig.1C).

Results



Results

Table 2 The IC₅₀ values (μM) of baicalein in breast cancer cells

Cell lines	24 hours	48 hours	72 hours
MCF-7	51.06	20.12	13.98
MDA-MB-231	60.12	27.96	19.01

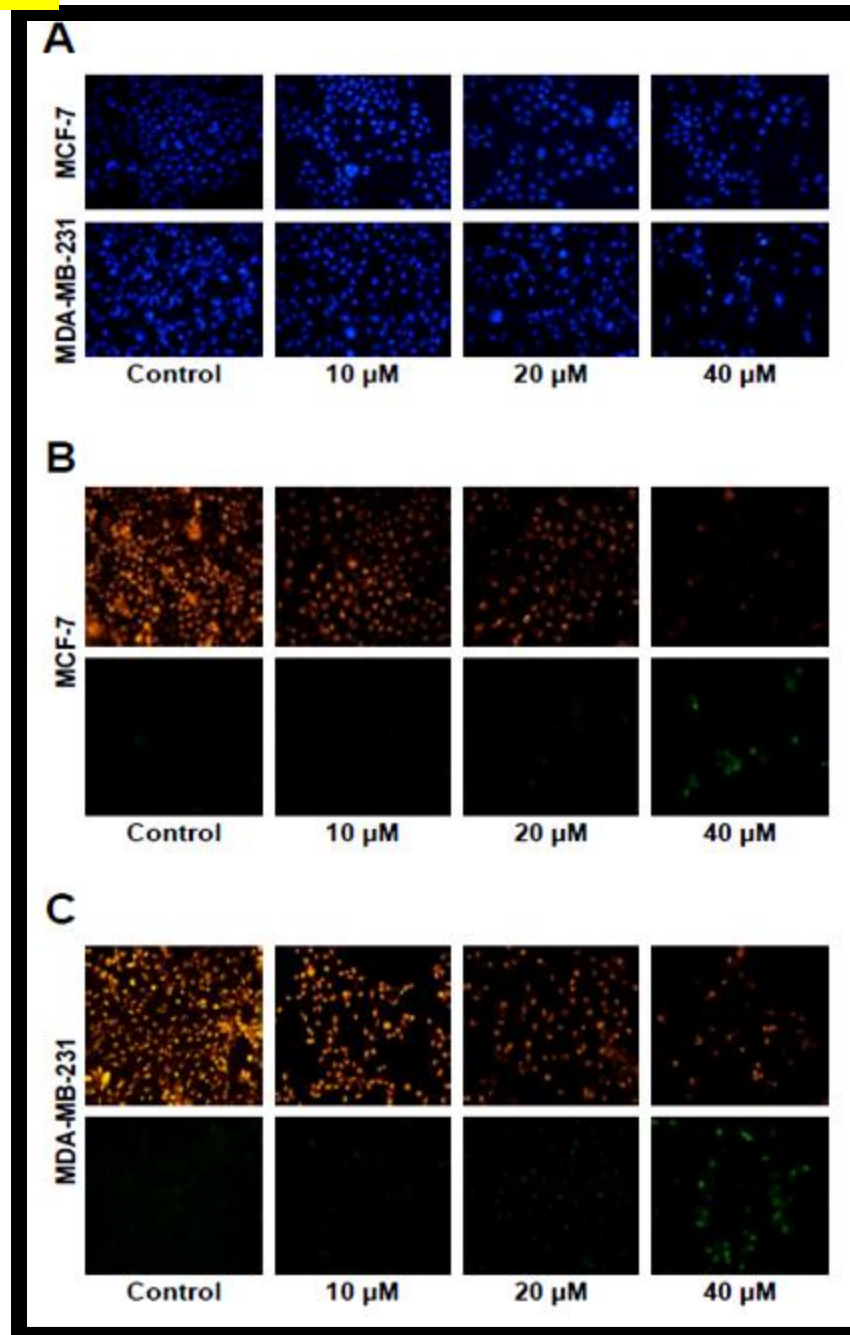
Abbreviation: IC₅₀, 50% inhibition concentration.

Results

Baicalein induced cell **apoptosis** in breast cancer cells

- ◉ **Hoechst staining 33258** (sensitive to DNA & used to assess changes in cellular nuclear morphology), baicalein effects on nuclear condensation (Fig.2A).
- ◉ **mitochondrial membrane potential ($\Delta\Psi_m$)** changes (key events during drug-induced apoptosis were examined via fluorescent probe JC-1).
- ◉ normal cells: JC-1 accumulates & aggregates which characterized by red fluorescence in the mitochondria.
- ◉ During apoptosis: $\Delta\Psi_m$ changes → JC-1 aggregates transform into monomers leading to loss of red fluorescence & $\Delta\Psi_m$ changes are manifested via reduced ratio of red to green fluorescence.

Results



Results

- ❖ **Annexin V-FITC/PI staining and flow cytometry:**

- ⊙ **Baicalein increases cell apoptosis in MCF-7 and MDA-MB-231 cells with an apoptotic rate estimated at:**

**26.89%±0.96% and 27.73%±0.23% in 40 μM baicalein group,
20.55%±0.62% and 20.27%±0.36% in 20 μM baicalein group.
13.08%±0.78% and 16.94%±0.86% in 10 μM baicalein group.
and 8.62%±0.34% and 6.89%±0.65% in the control group.**

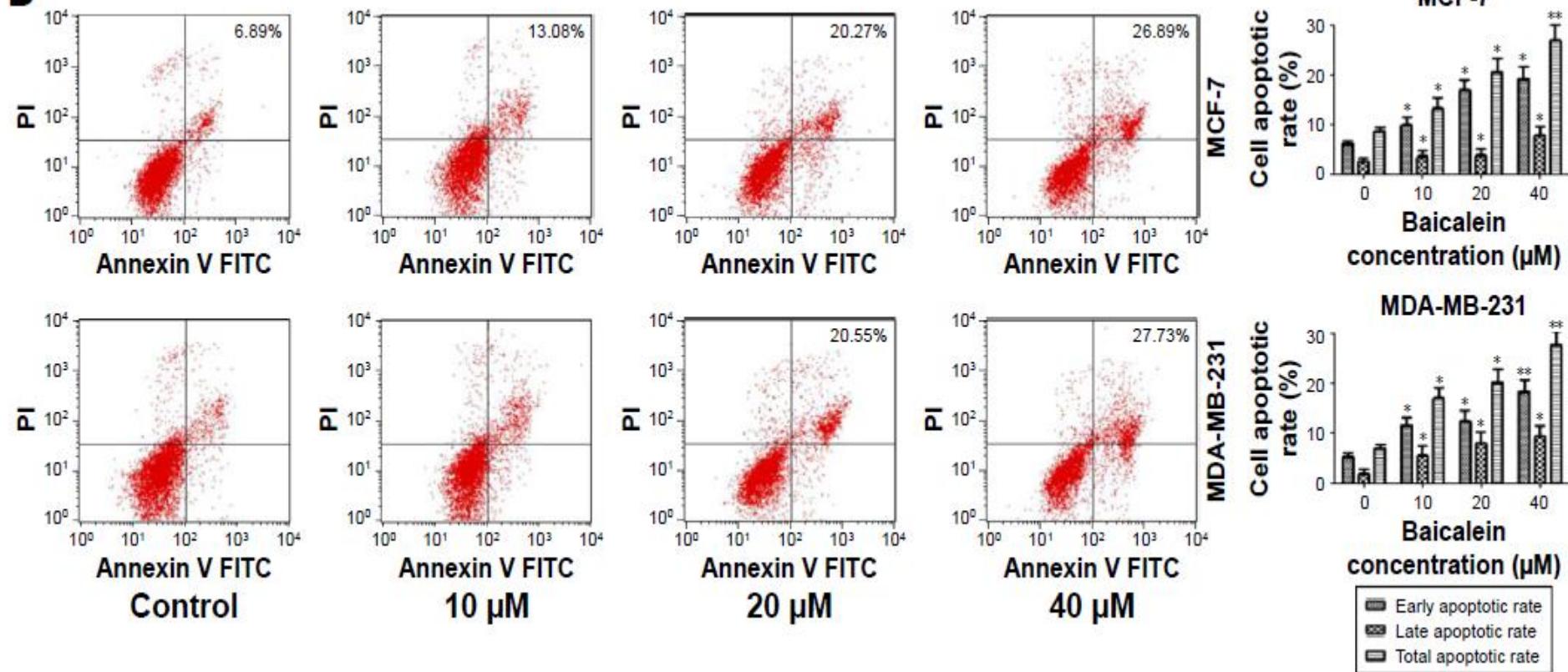
(Figure 2D).

- ❖ **qRT-PCR and Western blotting: the expression of apoptosis-related genes and proteins (Fig.2E & F):**

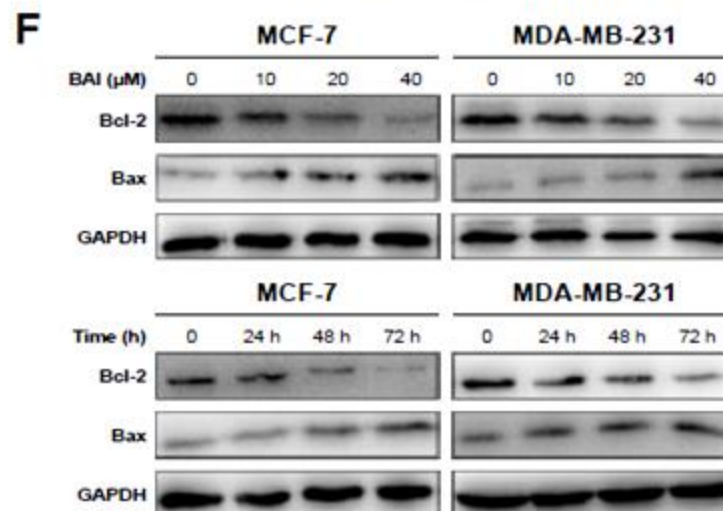
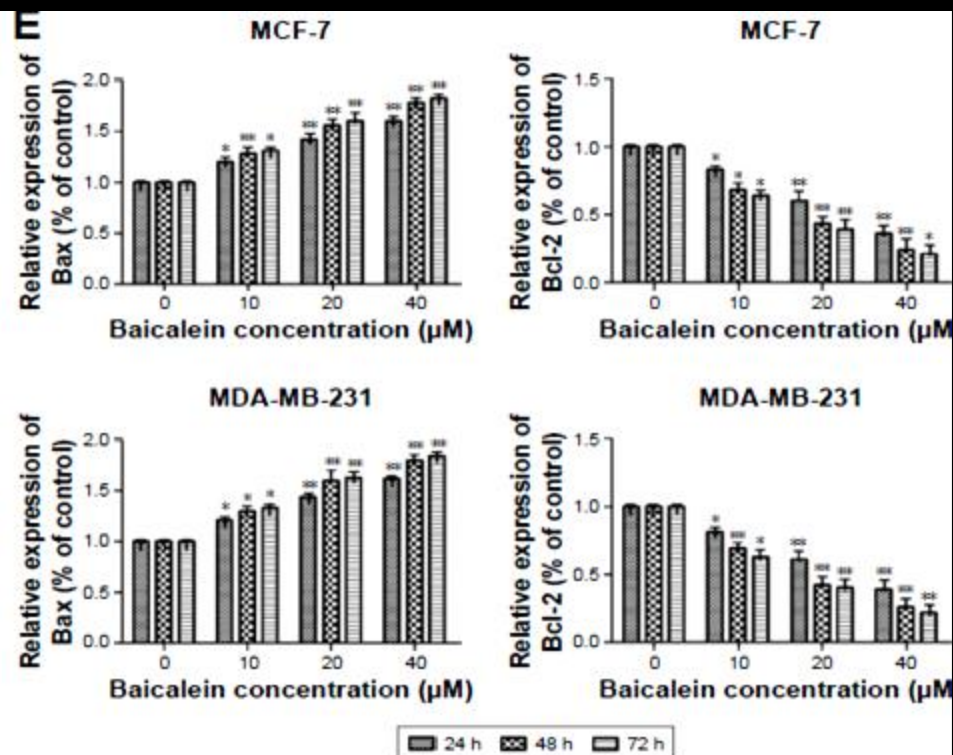
- ⊙ **↑ Bax & ↓ Bcl-2.**

Results

D



Results

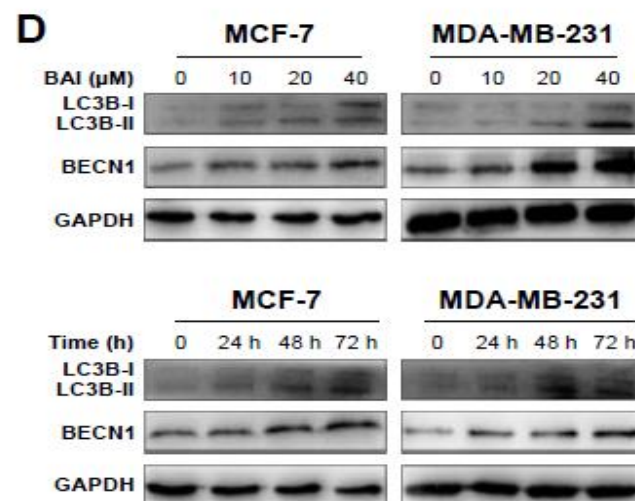
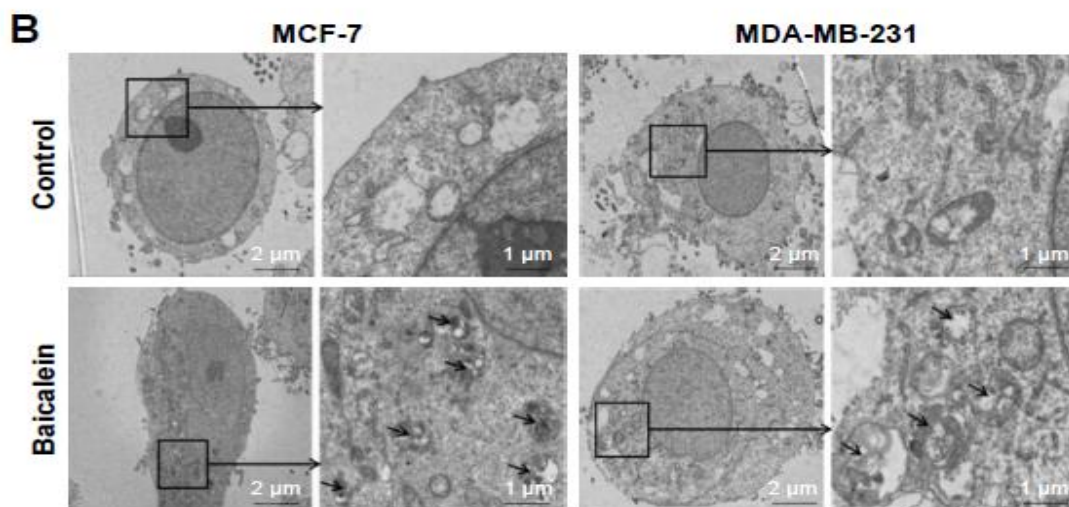
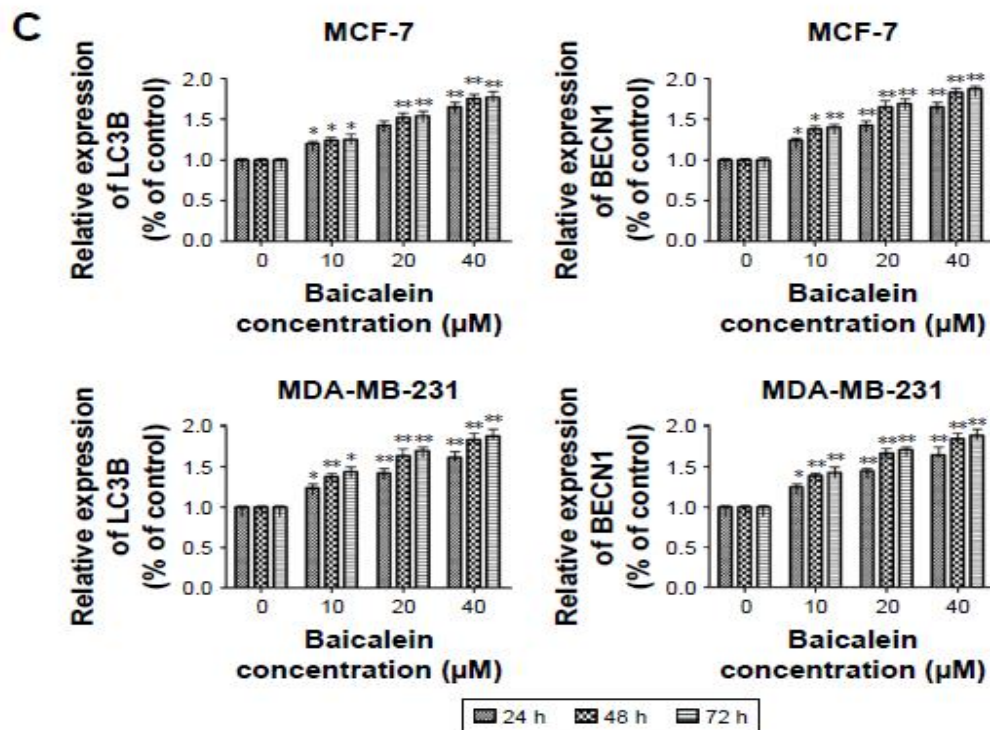
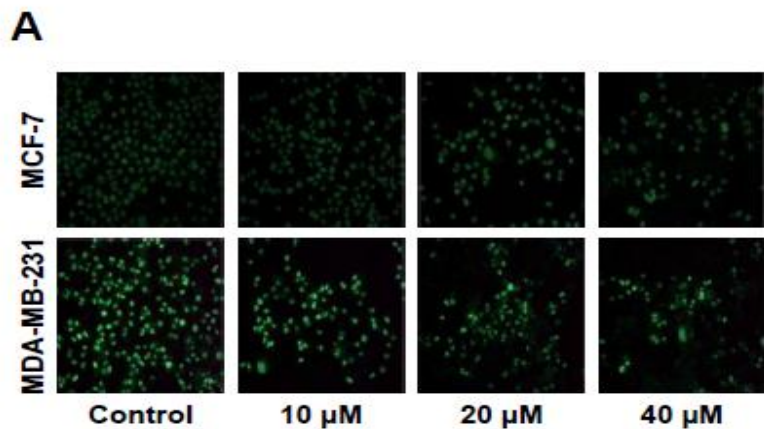


Results

Baicalein induces **autophagy in MCF-7 and MDA-MB-231 cells**

- ⊙ **Baicalein induced the formation of autophagic vacuoles in MCF-7 and MDA-MB-231 cells in a dose-dependent manner (Fig.3A).**
- ⊙ **Changes in morphological and ultrastructural features of cells were visualized. (Cells were treated with 40 μ M baicalein for 48 hours) (Figure 3B).**
- ⊙ **To further confirm the autophagy induced by baicalein, the expression of autophagy-related genes and proteins (LC3B and BECN1) was detected by qRT-PCR and Western blotting (Fig.3C and D).**

Results



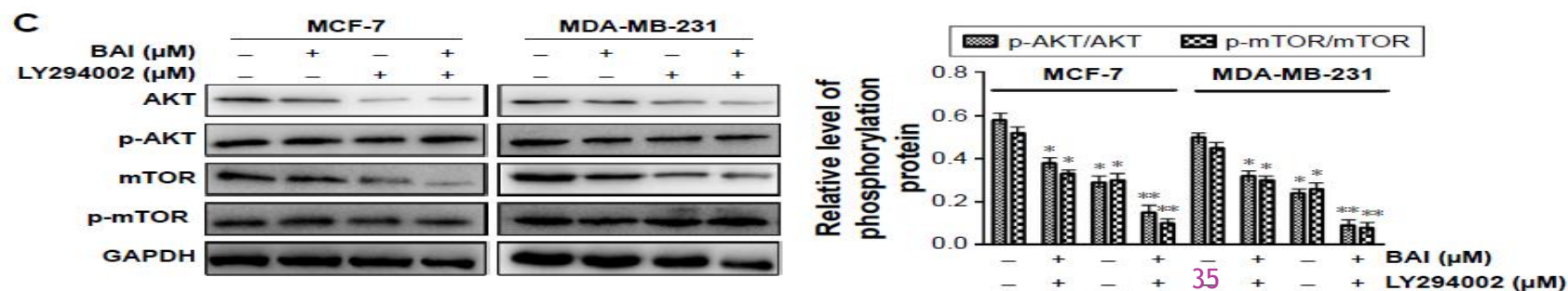
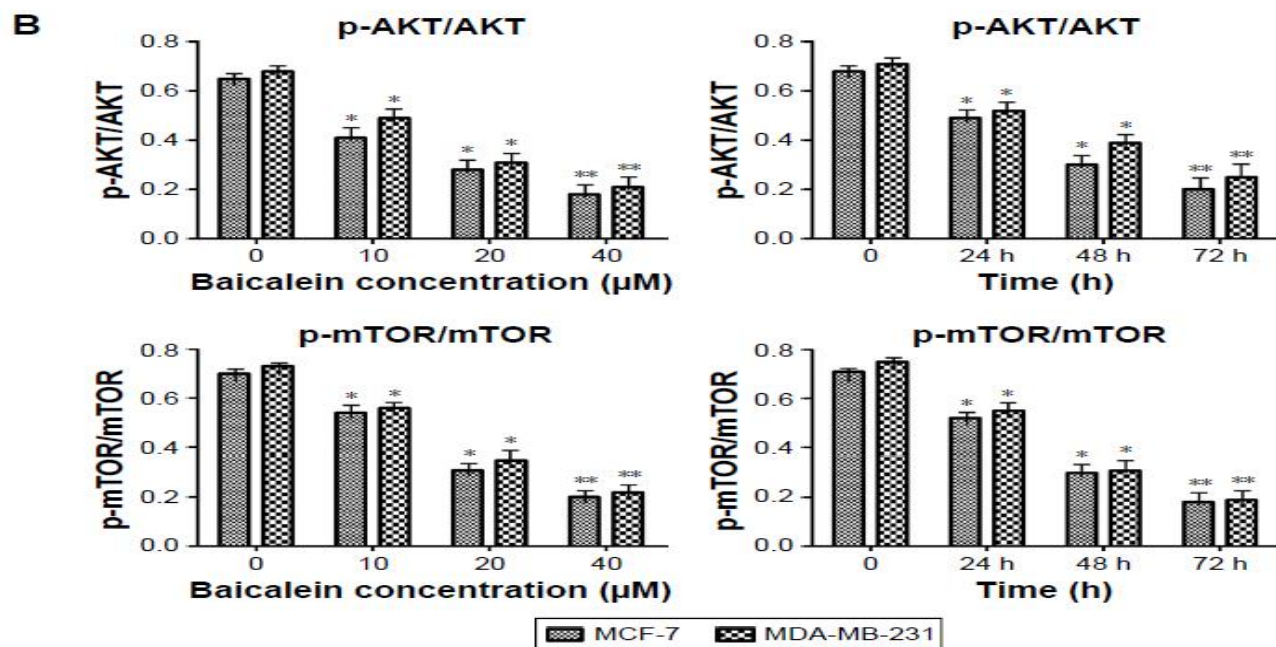
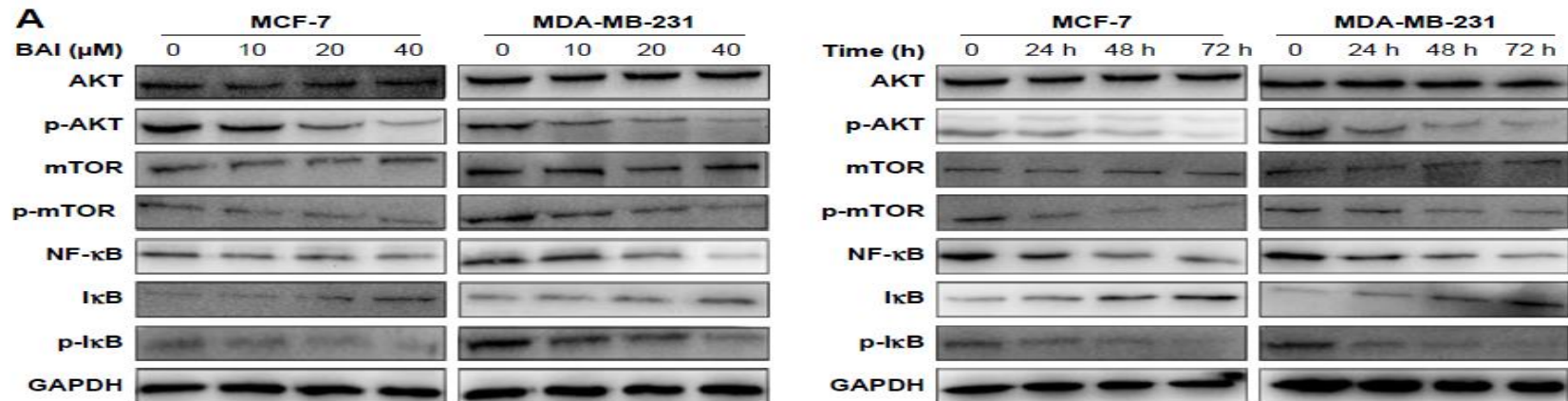
Results

Baicalein regulates **the apoptosis and autophagy** of breast cancer cells via the **PI3K/AKT/AKT signaling** pathway

- ❖ To elucidate the molecular mechanism of baicalein-induced apoptosis & autophagy, levels of proteins associated with the PI3K/AKT signaling pathway were examined using Western blotting:

p-AKT, p-mTOR, NF- κ B, & p-I κ B protein expression was 
p-AKT/AKT & p-mTOR/mTOR ratios  (Fig.4B).

- ❖ To further confirm of the baicalein effects on PI3K/AKT signaling pathway: Breast cancer cells were treated with LY294002 (a specific PI3K inhibitor) \rightarrow p-AKT & p-mTOR reduction (Fig.4C).
- ⦿ These findings support the hypothesis that the induction of apoptosis and autophagy in cells by baicalein is mediated by the suppression of the PI3K/AKT signaling pathway.

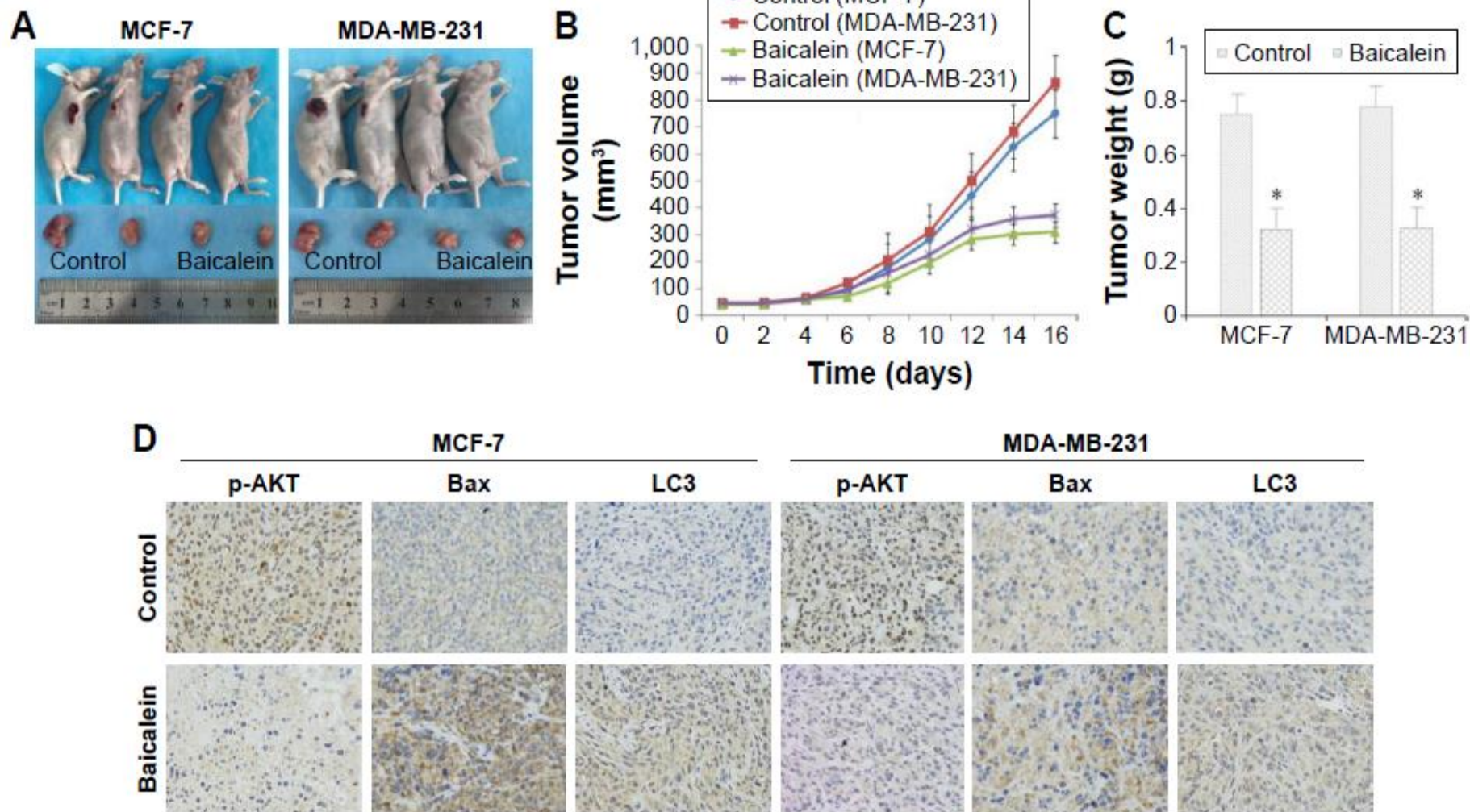


Results

In vivo effects of baicalein on breast cancer xenograft model

- ◉ In vitro: baicalein can inhibit proliferation and induce apoptosis and autophagy.
- ◉ In vivo: anticancer potential of baicalein were examined using breast xenografts models (BALB/c-nude).
- ◉ Treatment with baicalein → tumor tissues were collected and analyzed →
↓ **growth**, **volume**, and **weight** of tumors in the baicalein-treated group (Fig.5A–C)
- ❖ **Immunohistochemistry staining:** baicalein: ↓ p-AKT expression &
↑ expression of Bax & LC3 at the Pr level (Fig.5D) → baicalein can induce apoptosis & autophagy via modulation of the PI3K/AKT pathway.

Results



Discussion

- ⊙ **Previous in vivo & in vitro studies:** baicalein induces apoptosis in lung cancer, cervical cancer, esophageal carcinoma and Burkitt lymphoma cells.
- ⊙ There was a lack of information about the anticancer activity of baicalein in breast cancer, & on the signaling pathways related to apoptosis & autophagy.
- ❖ **In present study established that baicalein:**
 - ⊙ Inhibited cell proliferation in a time- and dose-dependent manner in MCF-7 & MDA-MB-231 cells by MTT and clone formation assays.
 - ⊙ Induced apoptosis in 2 breast cancer cell lines via Hoechst staining, $\Delta\Psi_m$, and annexin V-FITC assay.
 - ⊙ Induced apoptosis by increasing the Bax/BCL-2 ratio, as measured by qRT-PCR and Western blotting.

Discussion

- ❖ **baicalein-induced autophagy assessment:**
- ⊙ **The expression of BECN1 and LC3 was increased as shown by qRT-PCR and Western blotting.**
(BECN1 and LC3, a central protein and the initiator of autophagy, are regarded as autophagy-related proteins (autophagosome formation & maturation)).
- ⊙ **Autophagic cells were analyzed through acidic vesicular organelle staining assay, & autophagosomes detection under TEM.**

Discussion

- ❖ **The PI3K/AKT signaling pathway plays:**
 - ⊙ **A crucial role in regulating normal cell proliferation, differentiation, and apoptosis.**
 - ⊙ **Modulating the development & progression of cancers**
 - ⊙ **This signaling has been activated AKT, NF- κ B, and mTOR(which are the downstream components of the PI3K/AKT pathway) → when continuously activated → maintenance of malignancies.**
 - ⊙ **Western blot were applied to evaluate the expression level of this proteins (AKT, p-AKT, mTOR, p-mTOR, NF- κ B, I κ B, and p-I κ B) in the PI3K/AKT pathway.**

Discussion

- ⊙ **Results: baicalein reduced the expression of p-AKT, p-mTOR, NF- κ B.**
- ⊙ **These findings further supported the hypothesis that the induction of apoptosis and autophagy in cells by baicalein is mediated by the suppression of the PI3K/AKT pathway.**
- ⊙ **In addition, to acquire more reliable evidence to support and verify our in vitro experimental findings, we used the xenograft nude mouse model.**

Conclusion

Conclusion

- ◎ baicalein had the potential to:
- ◎ **suppress cell proliferation, induce apoptosis and autophagy** in MCF-7 and MDA-MB-231 breast cancer cells via inhibiting the PI3K/AKT pathway both in vitro & in vivo.
- ◎ These results suggest that baicalein may have **therapeutic potential** for breast cancer treatment.
- ◎ The anti-tumor function of baicalein has not been investigated in **clinical trials**, further study of the mechanisms that underpin baicalein's anti-tumor activity may provide possible clinical applications in the treatment of breast cancer.

A watercolor illustration featuring a central white rectangular area with the text "thank you" in a bold, black, cursive script. This central area is surrounded by a decorative border of watercolor-painted flowers and leaves. On the left side, there are clusters of bright yellow flowers and green leaves. On the right side, there are large, vibrant pink flowers with green foliage. The entire composition is set against a white background, which is itself framed by a thin yellow border. The overall style is soft and artistic, typical of watercolor painting.

thank
you